

## 175. Synthesis of 2-Acetamido-2-deoxy-D-gluconhydroximolactone- and Chitobionhydroximolactone-Derived *N*-Phenylcarbamates, Potential Inhibitors of $\beta$ -*N*-Acetylglucosaminidase

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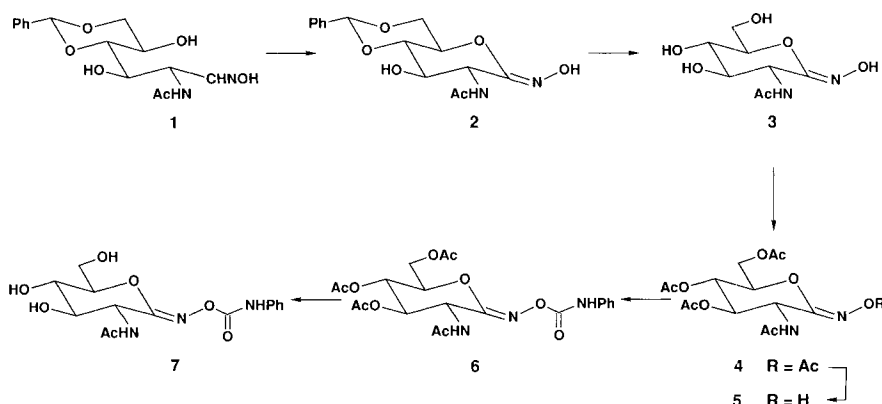
(10. VIII. 90)

The *N*-phenylcarbamate **7**, derived from 2-acetamido-2-deoxy-D-gluconhydroximo-1,5-lactone (**3**) and the analogous *N*-phenylcarbamate **14**, derived from chitobionhydroximo-1,5-lactone (**10**) have been prepared as potential inhibitors of  $\beta$ -*N*-acetylglucosaminidases. The unambiguous synthesis of the hydroximo-1,5-lactone **3** involves oxidation of the oxime **1**, followed by deprotection with Na/NH<sub>3</sub>.

**Introduction and Plan.** – We have described the synthesis of the *N*-phenylcarbamate derived from gluconhydroximo-1,5-lactone [1] and the strong inhibition of emulsin by this compound [2], which also inhibits phosphofructokinase b [3].

Consequently, the *N*-phenylcarbamate **7** (*Scheme 1*), derived from the 2-acetamido-2-deoxygluconhydroximolactone **4** [1] and its analogue **14** derived from chitobionhydroximolactone **10** (*cf. Scheme 2*) became of interest as potential inhibitors of  $\beta$ -*N*-acetylglucosaminidase (*EC 3.2.1.30*). The amorphous hydroximo-1,5-lactone **3** had been obtained by oxidation (MnO<sub>2</sub>) of *N*-acetyl-D-glucosamine oxime, together with the isomeric, crystalline hydroximo-1,4-lactone [1]. Of these products, the latter is easily obtained pure, while isolation of the former is cumbersome and not easily reproduced.

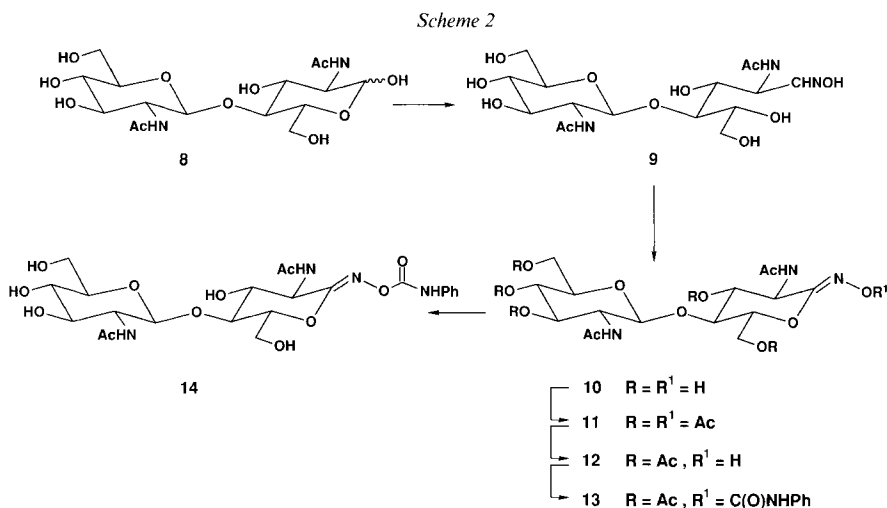
*Scheme 1*



We now report on a more convenient, if somewhat longer procedure for the preparation of the hydroximolactone **3**, on the preparation of the *N*-phenylcarbamate **7** from **3** via the selectively protected tetraacetyl derivative **5** [1], and on the analogous preparation of the *N*-phenylcarbamate **14** from chitobionhydroximolactone (**10**).

**Results.** – The partially protected oxime **1** (Scheme 1), available from *N*-acetyl-D-glucosamine in two steps and in a yield of 81% [1] [4], was oxidized with MnO<sub>2</sub> [5]. The crystalline hydroximo-1,5-lactone **2** was obtained in a yield of 50%, together with 35% of starting material. While catalytic debenzoylation of 2,3,4,6-Tetra-*O*-benzyl-D-gluconhydroximo-1,5-lactone had led to decomposition [6], debenzylidenation of **2** with Na/NH<sub>3</sub>, followed by chromatography on neutral alumina, afforded **3** in 86% yield. The pentaacetyl derivative **4**, obtained according to [1] was partially deacetylated (MeNH<sub>2</sub> in MeOH) to **5** (68% from **3**). This hydroximolactone reacted smoothly with phenyl isocyanate, and the resulting carbamate **6** was isolated in a yield of 94%. Transacetylation gave the desired, crystalline *N*-phenylcarbamate **7** (61%). It proved stable for many months when stored in the refrigerator.

To obtain the analogous *N*-phenylcarbamate, derived from chitobiose (**8**), we oxidized the oxime **9** (Scheme 2) with MnO<sub>2</sub> in the presence of a phosphate buffer at pH 7.



The hydroximolactone **10** was crystallized from MeOH (69%) and acetylated in Ac<sub>2</sub>O/pyridine to yield 83% of the hexaacetate **11**. Selective deacetylation (MeNH<sub>2</sub> in MeOH) gave the pentaacetyl derivative **12** (43%), and treatment of **12** with phenyl isocyanate afforded the protected *N*-phenylcarbamate **13** (76%). Transacetylation of **13** with NH<sub>3</sub> in MeOH gave 46% of the carbamate **14**. All these compounds, derived from chitobiose crystallize easily and have relatively high melting points.

The carbamate **7** is a strong inhibitor of β-*N*-acetylglucosaminidase. Its biochemical properties and those of the hydroximolactone **3** will be reported separately [7].

We thank *Sandoz Ltd.*, Basel, and the *Swiss National Science Foundation* for generous support.

## Experimental Part

*General.* Cf. [8]. All solvents were distilled before use; H<sub>2</sub>O was distilled twice. Activated MnO<sub>2</sub> was prepared following [5]. TLC: Merck precoated silica gel 60 F<sub>254</sub> plates; detection by spraying with a 0.025M I<sub>2</sub> soln. in 10% aq. H<sub>2</sub>SO<sub>4</sub>, followed by heating at ca. 200°. Column chromatography: silica gel Merck 60; flash chromatography (FC): 40–63 µ; medium-pressure chromatography (MPLC) [9]: 15–40 µ. Solvents mixtures: A = AcOEt/MeOH/H<sub>2</sub>O, B = CHCl<sub>3</sub>/EtOH. M.p.: not corrected. Optical rotations: Perkin-Elmer-241 polarimeter, 1-dm cell, measured at 365, 436, 546, 578, and 589 nm; the specific rotation at 589 nm was determined using a regression curve. IR: 3% CHCl<sub>3</sub> solns., unless stated otherwise.

**2-Acetamido-4,6-O-benzylidene-2-deoxy-D-gluconhydroximo-1,5-lactone (2).** To a stirred soln. of **1** [**1**] (4 g, 12.3 mmol) in MeOH (150 ml) at 50° were added 2 equiv. of activated MnO<sub>2</sub> [5] (2.139 g, 24.6 mmol). The mixture was stirred overnight at 50° and filtered through Celite. Concentration of the filtrate to 40 ml followed by adsorption on silica gel and FC (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 10:1) afforded **1** (1.4 g, 35%) and **2** (1.98 g, 50%), which was crystallized from AcOEt/MeOH. *R*<sub>f</sub> (CH<sub>2</sub>Cl<sub>2</sub>/MeOH 8:1) 0.4. M.p. 229–231° (dec.).  $[\alpha]_D^{25} = +48.2$  (*c* = 0.2, MeOH). IR (KBr): 3360s (br.), 3095w, 3060w, 2870w, 1710w, 1660s (sh), 1645s (br.), 1565m (sh), 1550m, 1470w, 1450m, 1430w, 1390m, 1380m, 1330w, 1315m, 1250m, 1210w, 1150w, 1135m, 1120m, 1095s, 1045m, 1030m, 1000m, 970m, 945m, 925w, 895w, 875w, 790w, 760m, 750w, 700m. <sup>1</sup>H-NMR (400 MHz, (D<sub>6</sub>)DMSO): 10.08 (s, NOH); 8.26 (*d*, *J* = 8.4, NH); 7.46–7.43 (*m*, 2 arom. H); 7.38–7.36 (*m*, 3 arom. H); 5.68 (*s*, HO–C(3), PhCH); 4.36–4.34 (*m*, 2 H); 3.85–3.7 (*m*, 4 H); 1.85 (*s*, CH<sub>3</sub>). <sup>13</sup>C-NMR (50 MHz, (D<sub>6</sub>)DMSO): 169.1 (*s*, CH<sub>3</sub>CO); 150.25 (*s*, C(1)); 137.6 (*s*, arom. C); 129.2 (*d*, arom. C); 128.3 (*2d*, arom. C); 126.5 (*2d*, arom. C); 100.9 (*d*, PhCH); 80.4 (*d*, C(4)); 71.3 (*d*); 68.4 (*d*), 67.6 (*t*, C(6)); 52.5 (*d*, C(2)); 23 (*q*, CH<sub>3</sub>). CI-MS (NH<sub>3</sub>): 340 (54, [M + 1 + NH<sub>3</sub>]<sup>+</sup>), 323.3 (25, [M + 1]<sup>+</sup>). Anal. calc. for C<sub>15</sub>H<sub>18</sub>N<sub>2</sub>O<sub>6</sub> (322.32): C 55.90, H 5.63, N 8.69; found: C 55.77, H 5.85, N 8.49.

**2-Acetamido-2-deoxy-D-gluconhydroximo-1,5-lactone (3).** Five equiv. of Na (759 mg, 33 mmol) were added in portions at –35° to a soln. of **2** (2 g, 6.6 mmol) in NH<sub>3</sub> (45 ml), until the blue color persisted for more than 1 min, and TLC showed the disappearance of **2**. Evaporation of NH<sub>3</sub> left a residue which was redissolved in MeOH and purified by FC (neutral alumina, AcOEt/MeOH/H<sub>2</sub>O 7:2:1): **3** (1.25 g, 86%). *Data of 3*: in agreement with those in [1].

**O-(2-Acetamido-2-deoxy-D-glucoopyranosylidene)amino N-Phenylcarbamate (7).** Phenyl isocyanate (290 µl, 2.64 mmol) and Et<sub>3</sub>N (950 µl, 6.6 mmol) were added a stirred soln. of **5** (480 mg, 1.32 mmol) [**1**] in THF (15 ml). After 3 h, the solvents were evaporated and the residue purified by FC (silica gel, AcOEt/hexane 3:2): **O-(2-acetamido-3,4,6-tri-O-acetyl-D-glucoopyranosylidene)amino N-phenylcarbamate (6, 600 mg, 94%)** as a foam. *R*<sub>f</sub> (AcOEt/hexane 5:2) 0.32. IR (CHCl<sub>3</sub>): 3424w, 3380w, 3320w (br.), 3030w, 3000w, 2960w, 1750s (br.), 1680m, 1395m (sh), 1370m, 1320w (sh), 1310m, 1210m, 1030m, 1020m, 990w, 895w. <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>): 7.82 (*s*, PhNH); 7.45–7.28 (*m*, 4 arom. H); 7.14 (*d*, *J* = 7.8, AcNH); 7.11 (*m*, arom. H); 5.45 (*dd* app. as *t*, *J* = 8.8, H–C(3)); 5.33 (*dd* app. as *t*, *J* = 8.7, H–C(4)); 4.85 (*dd* app. as *t*, *J* = 8.4, H–C(2)); 4.54 (*ddd* app. as *dt*, *J* = 2.8, 8.6, H–C(5)); 4.42 (*dd*, *J* = 3.2, 12.9, H–C(6)); 4.3 (*dd*, *J* = 2.5, 12.9, H–C(6)); 2.12 (*s*, CH<sub>3</sub>); 2.05 (*s*, CH<sub>3</sub>); 2.04 (*s*, CH<sub>3</sub>); 2.0 (*s*, CH<sub>3</sub>). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 170.5 (*s*, CO); 170.3 (*s*, CO); 170.2 (*s*, CO); 169.1 (*s*, CO); 155.2 (*s*, CONHPh); 151.8 (*s*, C(1)); 136.8 (*s*, arom. C); 129.2 (*2d*); 124.3 (*d*); 119.2 (*2d*); 77.2 (*d*); 71.3 (*d*); 67.1 (*d*); 61.2 (*t*, C(6)); 49.6 (*d*, C(2)); 22.8 (*q*, CH<sub>3</sub>); 20.62 (*q*, CH<sub>3</sub>); 20.57 (*q*, CH<sub>3</sub>); 20.5 (*q*, CH<sub>3</sub>).

Sat. NH<sub>3</sub>/MeOH (ca. 4M, 2 ml) was added to an ice-cold soln. of **6** (300 mg, 0.626 mmol) in MeOH (15 ml). After 4 h, the soln. was concentrated to 5 ml and purified by FC (AcOEt/MeOH 10:1) to afford **7** (135 mg, 61%) which was crystallized from AcOEt/MeOH/hexane. *R*<sub>f</sub> (AcOEt/MeOH 10:1) 0.21. M.p. 171–174° (dec.).  $[\alpha]_D^{25} = +64.8$  (*c* = 0.2, MeOH). IR (KBr): 3470m (sh), 3420s, 3310s, 3060w, 2950w, 2930w, 2900w, 2880w, 1745s, 1705m, 1660m, 1645s, 1605m, 1600m (sh), 1553m, 1530s, 1495w, 1450m, 1415w, 1380m, 1330w, 1310m, 1290w (sh), 1260w, 1205m, 1295m, 1160w, 1130w, 1005m, 1080m, 1070m, 1040m, 1020m, 1000w, 960w, 930w, 900w, 880w, 860w, 770w, 750m, 690m, 640m, 620m. <sup>1</sup>H-NMR (400 MHz, CD<sub>3</sub>OD): 7.44 (*m*, 2 arom. H); 7.29 (*m*, 2 arom. H); 7.05 (*m*, H arom.); 4.58 (*m*, X of ABX, H–C(2)); 3.95 (*dd*, *J* = 2.2, 12.8, H–C(6)); 3.94 (*m*, H–C(5)); 3.85 (*dd*, *J* = 4.6, 12.8, H–C(6)); 3.75 (*m*, AB of ABX, H–C(3), H–C(4)); 2.06 (*s*, CH<sub>3</sub>). <sup>13</sup>C-NMR (50 MHz, (D<sub>6</sub>)DMSO): 169.3 (*s*, CH<sub>3</sub>CO); 158.2 (*s*, CONHPh); 151.8 (*s*, C(1)); 138.6 (*s*); 128.8 (*2d*); 122.9 (*d*); 118.7 (*2d*); 82.4 (*d*); 72.3 (*d*); 68.6 (*d*); 60 (*t*, C(6)); 51 (*d*, C(2)); 22.8 (*q*, CH<sub>3</sub>). Anal. calc. for C<sub>15</sub>H<sub>19</sub>N<sub>3</sub>O<sub>7</sub> (353.33): C 50.99, H 5.42, N 11.89; found: C 51.08, H 5.66, N 11.6.

**Chitobiose (= 2-Acetamido-4-O-(2-acetamido-2-deoxy-β-D-glucoopyranosyl)-2-deoxy-D-glucoopyranose; 8).** To a soln. (pH ~ 3) of a dried, crude hydrolysis product (2.84 g<sup>1)</sup> of chitin in MeOH was added 2.3 g of NaOAc. Ac<sub>2</sub>O (14 ml) was added dropwise over 30 min to the neutral mixture at r.t. The mixture was concentrated *i.v.* MPLC (300 g; A 63:25:12) of the residue afforded 1.6 g of a colorless syrup, which crystallized spontaneously *i.v.*

<sup>1)</sup> We thank Dr. H. Braunschweiger, Sandoz AG, Basel, for a generous gift of the crude hydrolysate.

Recrystallization in EtOH afforded an anal. pure sample, whose physical properties were identical with those of a commercial sample of chitobiose (*Sigma*, ref.-No. *D-1523*).

**Chitobiose Oxime (9).** A hot soln. of NaOEt, prepared from Na (27.6 mg, 1.2 mmol) in EtOH (6 ml), was added to a vigorously stirred soln. of powdered hydroxylammonium chloride (90.3 mg, 1.3 mmol) in EtOH (11 ml). The mixture was cooled in an ice-bath and filtered. Then, **8** (424 mg, 1 mmol) was added with stirring (pH ~ 7) at 50°. After disappearance of **8** (TLC, *A* 65:23:12), the mixture was concentrated *i. v.* Recrystallization of the residue (434 mg, 97%) in hot EtOH gave a sample of **9** for analysis.  $R_f$  (*A* 65:23:12) 0.24.  $[\alpha]_D = -0.8$  ( $c = 1.09$ , H<sub>2</sub>O, final value). IR (KBr): 3370 (br.), 3100 (sh), 2970 $m$ , 2930 $m$ , 2890 (br.), 1740 $s$ , 1550 (br.), 1420 (br.), 1374 $s$ , 1315 $s$ , 1230 (sh), 1150 (sh), 1100 (sh), 1077 $s$ , 1039 (br.), 940 $m$ , 875 $m$ . FAB-MS: 440 ( $[M + 1]^+$ ).

**Chitobionhydroximo-1,5-lactone (10).** Activated MnO<sub>2</sub> [5] (800 mg) was added to a stirred soln. of **9** (980 mg, 2.2 mmol) in a mixture of KH<sub>2</sub>PO<sub>4</sub> (1.03 g) and NaOH (0.18 g) in H<sub>2</sub>O (60 ml; pH 7). The mixture was stirred at 40° for 5 h, until TLC (*A* 65:23:12) indicated the disappearance of **9**. The mixture was filtered through a pad of *Celite* and the filtrate was concentrated *i. v.* MPLC (80 g, *A* 65:23:12) of the residue gave a colorless syrup (673 mg, 69%), which crystallized spontaneously *i. v.* and was recrystallized in MeOH.  $R_f$  (*A* 7:2:1) 0.16. M.p. 244–246° (dec.).  $[\alpha]_D = +40.4$  ( $c = 1.04$ , H<sub>2</sub>O). IR (KBr): 3500 (sh), 3400 (br.), 3290 (sh), 3265 $s$ , 2920 $m$ , 2875 $m$ , 1660 $s$ , 1570 $m$ , 1550 $m$ , 1430 $m$ , 1400 (sh), 1372 $s$ , 1343 $m$ , 1307 $s$ , 1260 $m$ , 1250 $s$ , 1210 $m$ , 1160 $m$ , 1112 $s$ , 1090 $s$ , 1070 $s$ , 1050 $s$ , 1028 $s$ , 1010 $s$ , 995 $s$ , 940 $s$ , 875 $m$ , 752 $m$ . <sup>13</sup>C-NMR (50.4 MHz, D<sub>2</sub>O): 175.1 (*s*, CH<sub>3</sub>CO); 174.8 (*s*, CH<sub>3</sub>CO); 153.2 (*s*, C(1)); 101.7 (*d*, C(1')); 79.8 (*d*); 77.6 (*d*); 76.4 (*d*); 73.9 (*d*); 71.6 (*d*); 70.2 (*d*); 61.1 (*t*); 60.2 (*t*); 56.0 (*d*); 50.9 (*d*); 22.6 (*q*); 22.5 (*q*). FAB-MS: 438 ( $[M + 1]^+$ ). Anal. calc. for C<sub>16</sub>H<sub>27</sub>N<sub>3</sub>O<sub>11</sub> (437.402): C 43.94, H 6.22, N 9.61; found: C 44.17, H 6.16, N 9.37.

**O-(Chitobiosylidene)amino N-Phenylcarbamate (14).** Under cooling, Ac<sub>2</sub>O (15 ml) was added to a stirred suspension of **10** (656 mg, 1.5 mmol) in pyridine (50 ml). The mixture was warmed to 40° and stirred for 3.5 h, until TLC showed the disappearance of **10**. The mixture was concentrated under high vacuum and the residue worked up as usual (CHCl<sub>3</sub>/1M NaHCO<sub>3</sub>). Addition of CH<sub>2</sub>Cl<sub>2</sub>, Et<sub>2</sub>O, and hexane to the remaining syrup gave the *l*-N-3,3',4',6,6'-hexa-O-acetylchitobionhydroximo-1,5-lactone (**11**) as colorless crystals (858 mg, 83%).  $R_f$  (*B* 9:1) 0.41. M.p. 198–200° (dec.).  $[\alpha]_D = -20.1$  ( $c = 1.12$ , CHCl<sub>3</sub>). IR 3450 (sh), 3422 $w$ , 3300 (br.), 3030 (sh), 3000 $m$ , 2960 (sh), 1748 $s$ , 1680 $s$ , 1651 $m$ , 1648 $m$ , 1635 (sh), 1508 $s$ , 1425 (br.), 1368 $s$ , 1290 (sh), 1220 (br.), 1160 (sh), 1115 (br.), 1070 (sh), 1045 $s$ , 1000 $m$ . <sup>13</sup>C-NMR (50.4 MHz, CDCl<sub>3</sub>): 170.9, 170.9, 170.6, 170.5, 170.2, 169.5, 169.2, 168.1 (8 $s$ , CH<sub>3</sub>CO); 156.7 (*s*, C(1)); 100.5 (*d*, C(1')); 78.1 (*d*); 73.0 (*d*); 72.1 (*d*); 71.9 (*d*); 70.6 (*d*); 68.2 (*d*); 62.3 (*t*); 61.7 (*t*); 54.8 (*d*); 47.7 (*d*); 23.2, 22.9, 20.7, 20.6, 20.6, 20.5, 19.3 (8 $q$ , CH<sub>3</sub>CO).

To an ice-cold soln. of **11** (592 mg, 0.86 mmol) in MeOH (30 ml) was added dropwise over 50 min a MeNH<sub>2</sub> soln. (0.08M in EtOH, 10.7 ml). The mixture was immediately concentrated at r. t. and purified by FC (50 g, *B* 9:1): colorless crystals (239 mg, 43%) which were recrystallized in CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O to give 3,3',4',6,6'-penta-O-acetylchitobionhydroximo-1,5-lactone (**12**).  $R_f$  (*B* 9:1) 0.26. M.p. 192–193°. IR: 3660 $w$ , 3570 $m$ , 3425 $m$ , 3360 (br.), 3210 (sh), 3020 (sh), 3000 $m$ , 2970 (sh), 1742 $s$ , 1670 $s$ , 1510 $s$ , 1430 $m$ , 1370 $s$ , 1290 $m$ , 1220 $s$ , 1260 (br.), 1110 (sh), 1070 (sh), 1045 (sh), 1015 (sh), 945 $m$ .

Et<sub>3</sub>N (112  $\mu$ l) and phenyl isocyanate (305 mg) were added to a stirred soln. of **12** (260 mg, 0.40 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 ml) and THF (10 ml). After 2 min, TLC (*B* 9:1) showed complete reaction. The mixture was evaporated and purified by FC (35 g, *B* 95:5), yielding *O*-(3,3',4',6,6'-penta-O-acetylchitobiosylidene)amino *N*-phenylcarbamate (**13**) as colorless crystals (235 mg, 76%), which were recrystallized in CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O.  $R_f$  (*B* 9:1) 0.56. M.p. 129–131° (dec.). IR: 3680 $m$ , 3460 $m$ , 3380 $m$ , 3340 (br.), 3020 (sh), 3000 $m$ , 1750 $s$ , 1677 $s$ , 1600 (br.), 1520 $s$ , 1445 $s$ , 1370 $s$ , 1320 (sh), 1310 $m$ , 1295 $m$ , 1120 $m$ , 1075 $m$ , 1048 $s$ , 1015 (sh), 995 $m$ .

MeOH sat. with NH<sub>3</sub> (15 ml, ca. 4M) was added to an ice-cold soln. of **13** (198 mg, 0.26 mmol) in MeOH (30 ml). After 3 h, the mixture was taken to dryness. The remaining syrup was treated with MeOH (2 ml) to give 44 mg (31%) of crystals. Evaporation of the mother liquor and FC (9 g, *A* 78:15:7) gave another 21 mg (15%) of **14**.  $R_f$  (*A* 7:2:1) 0.34. M.p. 218–220°.  $[\alpha]_D = +28.1$  ( $c = 0.7$ ; H<sub>2</sub>O). IR (KBr): 3460 $s$ , 3300 $s$ , 3080 (sh), 2915 $m$ , 2840 $m$ , 2825 (sh), 1755 $s$ , 1740 $s$ , 1715 (br.), 1655 $s$ , 1640 $s$ , 1601 $s$ , 1560 (sh), 1550 (br.), 1525 $s$ , 1445 $m$ , 1370 $m$ , 1320 $m$ , 1300 $m$ , 1255 (br.), 1210 $m$ , 1160 (br.), 1105 $m$ , 1072 $s$ , 1035 (sh), 1020 $m$ . <sup>1</sup>H-NMR (400 MHz, D<sub>2</sub>O): 7.47–7.42 (*m*, 4 arom. H); 7.25 (1 arom. H); 4.80–4.66 (*m*, DHO); 4.25 (br., NH); 4.13–3.92 (*m*, 5 H); 3.85–3.73 (*m*, 4 H); 3.63–3.43 (*m*, 3 H); 2.12 (*s*, CH<sub>3</sub>CO); 2.08 (*s*, CH<sub>3</sub>CO). <sup>13</sup>C-NMR (100.6 MHz, D<sub>2</sub>O): 175.1, 174.9 (2 $s$ , CH<sub>3</sub>CON); 158.5 (*s*); 154.9 (*s*); 129.8, 125.6, 121.5 (3 $d$ , 5 arom. C); 101.7 (*d*, C(1')); 81.1 (*d*); 79.9 (*d*); 77.2 (*d*); 76.5 (*d*); 73.9 (*d*); 71.1 (*d*); 61.1, 60.3 (2 $t$ , C(6), C(6')); 56.1 (*d*), 50.9 (*d*); 22.5, 22.55 (2 $q$ , CH<sub>3</sub>CO). FAB-MS: 558 ( $[M + 1]^+$ ). Anal. calc. for C<sub>23</sub>H<sub>32</sub>N<sub>4</sub>O<sub>12</sub> (556.525): C 49.64, H 5.80, N 10.07; found: C 49.72, H 5.75, N 10.34.

## REFERENCES

- [1] D. Beer, A. Vasella, *Helv. Chim. Acta* **1985**, *68*, 2254.
- [2] D. Beer, A. Vasella, *Helv. Chim. Acta* **1986**, *69*, 267.
- [3] a) R. Barford, J. W. R. Schwabe, N. G. Oikonomakos, K. R. Acharya, J. Hajdu, A. C. Papageorgiou, J. L. Martin, J. C. A. Knott, A. Vasella, L. N. Johnson, *Biochemistry* **1988**, *27*, 6733; b) A. C. Papageorgiou, N. G. Oikonomakos, D. D. Leonidas, B. Bernet, D. Beer, A. Vasella, *Biochem. J.*, accepted.
- [4] W. Roth, W. Pigman, *J. Am. Chem. Soc.* **1960**, *82*, 4608.
- [5] J. Attenburrow, A. F. B. Cameron, J. H. Chapman, R. M. Evas, B. A. Hems, A. B. A. Jansen, T. Walker, *J. Chem. Soc.* **1952**, 1094.
- [6] B. Aebischer, Ph. D. thesis, Fribourg, Switzerland, 1983.
- [7] a) M. Horsch, L. Hoesch, A. Vasella, D. M. Rast, submitted to *Eur. J. Biochem.*; b) D. M. Rast, R. Furter, M. Horsch, G. W. Gooday, submitted to *J. Gen. Microbiol.*
- [8] B. Aebischer, J. H. Bieri, R. Prewo, A. Vasella, *Helv. Chim. Acta* **1982**, *65*, 2251.
- [9] H. Loibner, G. Seidl, *Chromatographia* **1979**, *12*, 600.